Post-harvest Green Pea Pod Rot Caused by Sclerotinia sclerotiorum in Korea

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In June 2017, in Gangneung, Gangwon Province, South Korea, green pea pods exhibited post-harvest rot symptoms. The fungus was isolated from infected pea pods and cultured on potato dextrose agar for identification. The morphological characteristics were examined, sequences of the internal transcribed spacer region and the β-tubulin (βtub) gene were analyzed, and the pathogenicity was confirmed according to Koch’s postulates. The morphology, phylogenetic analysis, and pathogenicity tests confirmed that Sclerotinia sclerotiorum was the causal agent. This study reports the first case of post-harvest green pea pod rot caused by S. sclerotiorum in Korea.

Keywords: Green pea, Pod rot, Post-harvest, Sclerotinia sclerotiorum

Sclerotinia sclerotiorum is a devastating, necrotrophic, and cosmopolitan plant pathogen with a broad host range (Boland and Hall, 1994; Bolton et al., 2005). In Korea, 60 plant species have been reported as hosts of S. sclerotiorum (Kim et al., 2009). The optimal temperature range for Sclerotinia growth is from 15–21°C, and cool and moist conditions increase the incidence of field rot. Green peas have a short season and a limited shelf-life, so preservation ensures long-term availability. Growth can also occur in lower temperatures (0–4°C); therefore, green peas are also susceptible to fungal growth during transit and storage (Naito and Sugimoto, 1986; Willetts and Wong, 1980). This study identifies S. sclerotiorum as the causal agent of a post-harvest green pea pod rot based on morphological characteristics, phylogenetic analysis, and a pathogenicity test.

Green pea pods harvested from Gangneung, Gangwon Province, South Korea (June 2017) exhibited post-harvest rot symptoms (white and fuzzy mycelial growth with black sclerotia) after 5–7 days of refrigeration (4°C, 70% relative humidity) (Fig. 1A). The pathogen was isolated by excising the infected pod tissue and then surface-sterilized in 1% sodium hypochlorite (NaOCl) for one minute. The tissue was rinsed with sterilized distilled water (3×), and then plated on potato dextrose agar (PDA; Difco, Detroit, MI, USA). After 4 days of room temperature (RT; 20±2°C) incubation, the culture was purified by the hyphal-tip method and maintained on PDA plates. Five morphologically similar fungal isolates were obtained from five green pea pod rot samples, including isolate GPSS003, which was further examined. The fungus was incubated on PDA medium at RT for 7 days and produced white-gray colonies, and after 3 weeks, showed small sclerotia on the plate peripheries (Fig. 1C). The sclerotia
were black, globosely, cylindrically, or irregularly shaped and between 1.0–7.3×1.2–5.2 mm in size (n=20).

Sclerotial germination was investigated by placing 10–15, 21-day-old sclerotia in sand Petri dishes. The sclerotia were washed, dried, and pressed into the sand (~2 mm depth). The dishes were incubated at RT with alternating 12-hr fluorescent light cycles and were watered three times per week with sterilized water until apothecia developed (Huang and Kozub, 1989; Smith and Boland, 1989). A dissecting microscope was used to cut the mature apothecia into thin slices, and the asci and ascospores were examined and photographed with a compound microscope (40× magnification). Sclerotia sand germination generated apothecia after 5–6 weeks of incubation at RT (Fig. 1D). The apothecia were cup-shaped with a yellowish-brown color and measured 0.6–1.8 cm (n=4). Asci from the apothecia were cylindrical and had eight spores ranging from 101.2–152.8×5.1–8.5 μm in size (n=10) (Fig. 1E). The ascospores were uniseriate, single-celled, hyaline, and ellipsoid, and between 8.2–13.6×4.0–5.8 μm in size (n=30) (Fig. 1F). These characteristics are consistent with the description of *Sclerotinia sclerotiorum* (Lib.) de Bary (Chang and Kim, 2003; Kohn, 1979) and summarized in Table 1. A representative isolate (GPSS003) was deposited in the Korean Agricultural Culture Collection, National Institute of Agricultural Science, Rural Development Administration, Wanjoo, South Korea (KACC48672), for future studies.

For molecular identification, the GPSS003 *S. sclerotiorum* isolate was grown in potato dextrose broth for 5 days at RT in a shaking incubator (150 rpm). The mycelia were filtered through Whatman No. 1 filter paper, and genomic DNA

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**Fig. 1.** (A) Rotting post-harvest green peas indicated by the fuzzy growths of mycelium and black sclerotia on the pod. (B) Green peas inoculated with *Sclerotinia sclerotiorum* developed pod rot symptoms after 5 days. (C) *S. sclerotiorum* three-week-old colonies and black sclerotia growing on potato dextrose agar medium. (D) Apothecia (arrows). (E) Ascus containing eight ascospores. (F) Ascospores.
was extracted using the DNeasy Plant Mini Kit (Qiagen Inc., Valencia, CA, USA). The DNA sequences of the ribosomal DNA internal transcribed spacers (ITS) region (ITS-5.8S rDNA) and beta-tubulin gene (\(\beta\text{tub}\)) were amplified by polymerase chain reaction (PCR) on a Mastercycler Gradient (Eppendorf, Hamburg, Germany) using primers for the ITS region (ITS1, 5'-TCCGTAGGTGAACCTGCGG-3'; ITS4, 5'-TCCTCCGCTTATGGATATGC-3'); and the \(\beta\text{tub}\) gene (Bt1a, 5'-TTCCCCGGTCTCACTTCTCATG-3'; Bt1b, 5'-GACGAGATCGTTCATGTTGAACTC-3') (Glass and Donaldson, 1995; White et al., 1990). The PCR cycling conditions for the ITS region were 94°C for 4 min and 35 cycles of 94°C (35 sec), 52°C (55 sec), 72°C (1 min), and 72°C for 10 min. The PCR cycling conditions for the \(\beta\text{tub}\) gene were 98°C for 3 min and 30 cycles of 98°C (10 sec), 67°C (15 sec), 72°C (2 min), and 72°C for 10 min. The PCR reactions contained 0.5 μl of each primer, 0.5 μl of Taq DNA polymerase (Bioneer, Daejeon, Korea), 0.5 μl of each dNTP, 2.5 μl of 10× PCR reaction buffer, 18.5 μl of distilled water, and 2.0 μl of template DNA (total volume 25 μl). The DNA concentrations were estimated on a 1% agarose gel by comparing the PCR product band intensity with a 100 bp DNA ladder. DNA sequencing was performed by SolGent Com. Ltd. (Daejeon, Korea), and the nucleotide sequences were searched using the GenBank database BLASTn tool (http://www.ncbi.nlm.nih.gov/BLAST/). Phylogenetic analysis was performed in MEGA7 by the neighbor-joining method (Kumar et al., 2016; Saitou and Nei, 1987).

BLAST analysis returned a 571 bp ITS-5.8S rDNA sequence and a 463 bp \(\beta\text{tub}\) sequence. The fungus identity was confirmed; the analysis showed 100% similarity to the S. sclerotiorum isolate CXL14041906 ITS-5.8S rDNA sequence (GenBank accession no. KX781301) and 100% similarity to the S. sclerotiorum SS1 isolate KF545202 \(\beta\text{tub}\) sequence. The representative isolate (GPSS003) sequences were deposited in the NCBI database (GenBank accession no. MG931017 for ITS-5.8SrDNA; accession no. MG931018 for \(\beta\text{tub}\)). The phylogenetic tree results (based on the combined ITS-5.8SrDNA and \(\beta\text{tub}\) sequences) placed the representative isolate within a clade that includes the S. sclerotiorum reference isolates (Fig. 2).

To determine fungal pathogenicity, six green pea pods were surface-sterilized (1% NaOCl), rinsed with sterile distilled water (3×), and dried (at RT). The sterilized pods were placed on moist filter paper in Petri dishes (20×20×5 cm). Three fungal mycelial discs (6.5 mm) were taken from the margins of actively growing 7-day-old colonies and placed into three sterilized pods. PDA discs were placed into three

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| Table 1. The mycological characteristics of the green pea isolated pathogen and Sclerotinia sclerotiorum |
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| Characteristic | Present isolate | S. sclerotiorum* |
| Colony | White-gray | White-gray |
| Color | Initially cushion-like, globular, or irregular; dark brown then black | Initially cushion-like or short-cylindrical; white then black |
| Sclerotium | 1.0–7.3 | - |
| Shape | 1.2–5.2 | - |
| Length (mm) | Cup-shaped with yellowish-brown coloring | Cup-shaped with yellowish-brown coloring |
| Width (mm) | 0.6–1.8 | 0.5–2.0 |
| Apothecium | Cylindrical, hyaline, 8-spored | Cylindrical, hyaline, 8-spored |
| Shape | 101.2–152.8 | 110–160.0 |
| Length (μm) | 5.1–8.5 | 6.0–10.0 |
| Width (μm) | Hyaline, ellipsoid to ovoid | Hyaline, ellipsoid to ovoid |
| Ascus | 8.2–13.6 | 9.0–14.0 |
| Shape | 4.0–5.8 | 4.0–6.0 |
| Ascospore | - | - |
| Length (μm) | - | - |
| Width (μm) | - | - |

*Described by Khon (1979).
Sterilized pods as a control. The Petri dishes were incubated in a growth chamber for 7 days at RT and 90±10% relative humidity (Han et al., 2013; Kim et al., 2014). White mycelia (a symptom of rot) were seen on the inoculated pea pods after 5 days. The fungal pathogen was re-isolated from the diseased lesions on the inoculated pods and exhibited the same morphological characteristics as the original isolates (Fig. 1B). Therefore, the fungal pathogen fulfilled Koch’s postulates, and S. sclerotiorum was identified as the post-harvest green pea pod rot causal agent.

Morphological characterization, pathogenicity test, and phylogenetic analysis of the ITS region and $\beta$-tub gene identified S. sclerotiorum as the pathogen isolated from the green peas. In Korea, green pea field rot from Sclerotinia infections has been documented, but the post-harvest disease has not (Chang and Kim, 2003; Kim et al., 2006). S. sclerotiorum-induced stem and pod field rot have been reported in, e.g., Canada (Farr and Rossman, 2021), Bangladesh (Islam et al., 2020), and Korea (Kim et al., 2006) but this is the first report of a post-harvest green pea pod rot in Korea. Post-harvest rotting caused by S. sclerotiorum can lead to complete product loss, making it an economically important issue. These findings suggest that Sclerotinia-induced rot may pose a threat to the long-term storage of produce, and an effective control strategy is needed.

Conflicts of Interest

No potential conflict of interest relevant to this article was reported.

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References


